

Methods for the Analysis of Microcystins in Water to sub-Parts Per Trillion Detection Levels

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Introduction



- Microcystins are cyclic peptides produced by <u>cyanobacteria</u>
- They are produced by overgrowth of algae, especially at higher water temperatures – Climate change, invasive species, global trade and agricultural practices can exacerbate the problem
- Microcystins can be toxic for plants and animals including humans
- Once ingested, microcystins travel to the liver, via the bile acid transport system, where most is stored. Some remains in the blood stream and may contaminate tissue. Microcystins bind covalently to protein phosphatases thus disrupting cellular control processes. Their <u>hepatotoxicity</u> may cause serious damage to the <u>liver</u>. Microcystins can strongly inhibit protein phosphatases type 1 (PP1) and 2A (PP2A), and are linked to <u>pansteatitis</u>.
- Over 80 toxic variants are known

Introduction



- Microcystin-containing 'blooms' are a problem in countries worldwide including China, Brazil, Australia, the USA and much of Europe
- WHO action limit = 1000 ng/L (1ppb) and methods are proposed including US EPA Methods 544 and 545 and European ISO 20179:2005(E) guideline
- For this reason sensitive detection is needed
- This presentation will discuss the various approaches to sample introduction and detection suitable for proposed regulatory limits as well as the lower limits often desired for research purposes

Experimental: UPLC conditions



- Run Time: 7.50 min
- Inj Vol: 50.00 μL
- Column: ACQUITY UPLC BEH C18 2.1x100mm, 1.7 μm
- Solvent A: 0.1% FA in 97/3 H2O/ACN
- Solvent B: ACN
- Flow Rate: 400 µL/min
- Gradient:

Time (min)		%A	%B	Curve
0.00	100.0	0.0	Initial	
0.75	100.0	0.0	6	
5.00	20.0	80.0	6	
6.00	0.0	100.0	1	
7.50	100.0	0.0	1	

Column Temp: 50.0 C

Experimental: MS Method Settings



- Time segmented MRM with Dwells to give 15 pts across each peak
- 2 MRMs per analyte
- Most microcystins singly charged except for DE-M-RR & M-RR

🗹 Experiment Setup - c:\masslynx projects\20160204 watergroep m	icrocystines.pro\acqudb\test_jd.exp	- • ×
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D 🖻 🖬 🎒 🗭 🗙		
😰 SIR 🛛 🖉 MRM 🛛 🕜 MS Scan 🛛 🗭 Parents 🖉 Daughters	📝 Neutral Loss 📝 Survey 📝 Phosphopeptide	
Points Per Peak: 17.467		
Total Run Time: 7.00 ↔	05mjins	
No. Type Information	Time	
2 MRM of 3 mass pairs, Time 0.00 to 3.00, ES+ (CYL)		
1 MRM of 4 mass pairs, Time 0.00 to 3.50, ES+ (ANA)		
3 MRM of 2 mass pairs, Time 2.00 to 5.00, ES+ (DE-RR)		
4 MRM of 2 mass pairs, Time 3.00 to 4.50, ES+ (M-RR)		
5 MRM of 3 mass pairs, Time 3.00 to 4.50, ES+ (NOD)		
10 MRM of 3 mass pairs, Time 3.00 to 4.50, ES+ (M-YR)		
7 MRM of 3 mass pairs, Time 3.00 to 5.00, ES+ (M-LR)		
8 MRM of 2 mass pairs, Time 3.50 to 5.50, ES+ (MC-LY)		
9 MRM of 3 mass pairs, Time 4.50 to 7.00, ES+ (M-LW)		
6 MRM of 3 mass pairs, Time 4.50 to 7.00, ES+ (M-LF)		
-		
		NUM

Performance Criteria

- The following performance criteria were evaluated
 - Linearity
 - Robustness
 - Sensitivity
 - Repeatability
 - Accuracy
 - Standard Addition



Xevo TQ-S shown with UPLC and APGC

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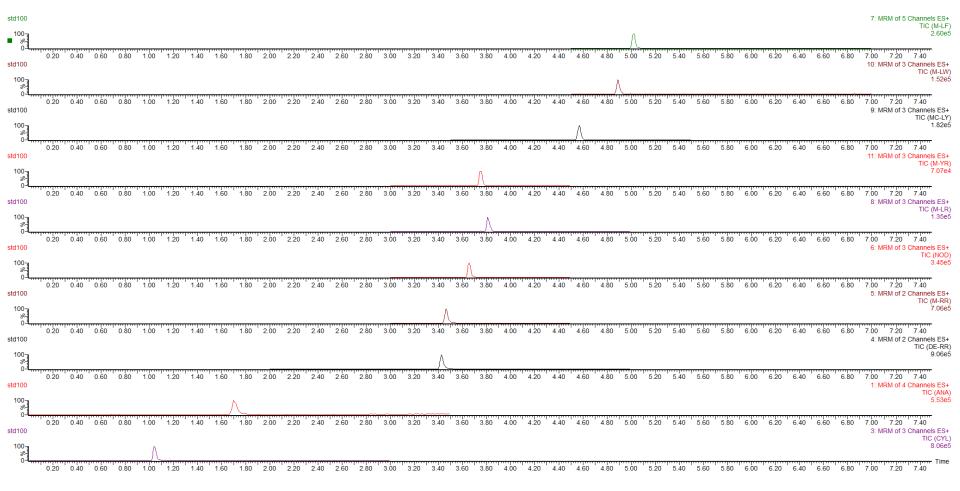
Mass Spectrometers SQ, TQ, QTof, & QTof with Ion Mobility ■

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Results: Typical Chromatogram

 Bottom to top: Cylindrospermopsin (CYL), Anatoxin-A (ANA), M-DE-RR, M-RR, Nodularin (NOD), M-LR, M-YR, M-LY, M-LW and M-LF at 100 ng/L



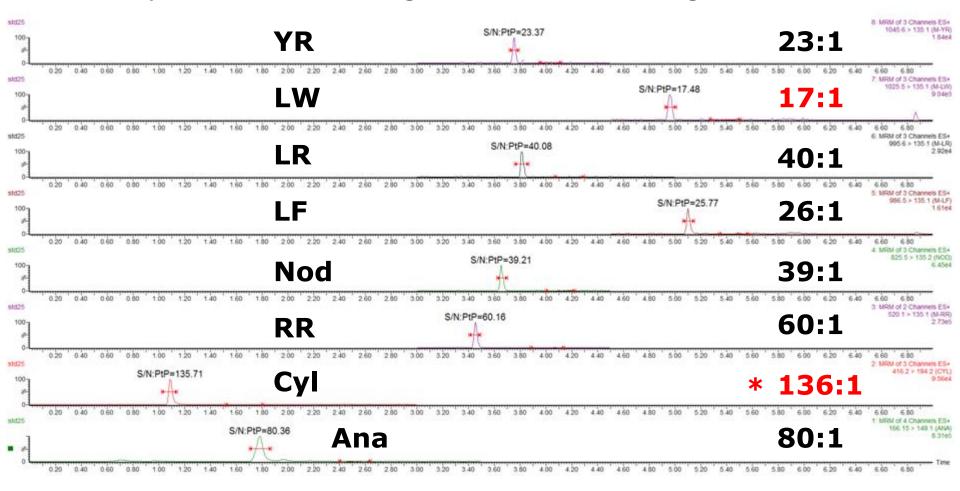
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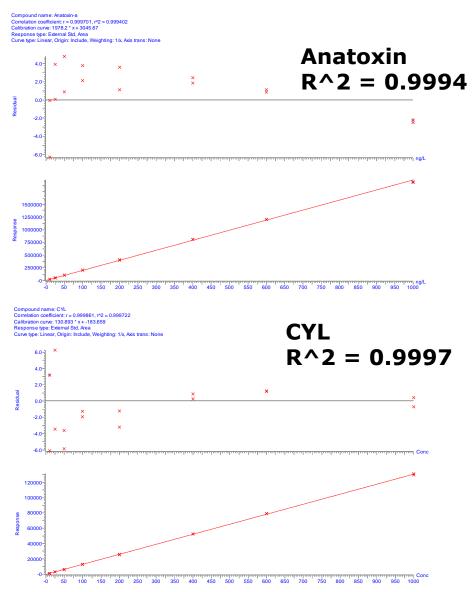


Results: Sensitivity

All analytes detected with good S:N at the 25 ng/L



Results: Linearity from 10 to 1000ng/L



Correlation coefficient: r = 0.999806, r^2 = 0.999613 Calibration curve: 70.7322 * x + 6.27616 Response type: External Std, Area Curve type: Linear, Origin: Include, Weighting: 1/x, Axis trans: None Nodularin 10.0-1 $R^2 = 0.9996$ 5.0 -5.0 -10.0 Compound name: MC-YR Correlation coefficient: r = 0.999714, r^2 = 0.999429 Calibration curve: 22.3124 * x + -22.0103 Response type: External Std, Area M-YR Curve type: Linear, Origin: Include, Weighting: 1/x, Axis trans: None $R^2 = 0.9994$ 4.0 2.0 0.0 -2.0 -4.0 -6.0 -8.0

Compound name: Nodularin

50 100

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m Cond

Results: Robustness



- Due to lack of IS, signal stability was evaluated
- First and last (5th) 100 ng/L std were compared. Time difference between samples approximately 15.5 hrs. Areas within 10% for 5 of the 8 compounds, and 31% for the other 3

Compound	100 ng/L (1)	100 ng/L (5) +15.5h	ratio
M-YR	2061	2215	107%
M-LW	1361	1660	122%
M-LR	2770	3353	121%
M-LF	1522	1990	131%
NOD	6848	7380	108%
M-RR	28133	29911	106%
CYL	12027	12074	100%
ANA	190364	183937	97%

Results: Repeatability



- Short term repeatability evaluated using 5 replicates from the same vial at 100 ng/L
- % CV values for DW and surface water SW
- Repeatability is good with % CV values below 5%

Compound	% CV	(n=5)
	DW	SW
		2.0
M-YR	4.6	3.0
M-LW	3.2	2.2
M-LR	2.2	1.8
M-LF	4.2	4.6
NOD	1.9	3.0
M-RR	1.6	0.5
CYL	5.0	2.7
ANA	1.6	0.5

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Results: Accuracy

- Accuracy evaluated using spike/recovery experiments at 100, 200 and 400 ng/L in DW and SW
- DW recoveries are acceptable for compounds eluting in the middle of the run but too low for compounds eluting near the beginning and the end
- Suggests the need for IS to improve accuracy

% Spike Recovery Values

Name	DW 1	DW 2 [DW 3	DW 4	DW 5
CYL	-54.7	-61.2	-59.6	-43.6	-40.8
Anatoxin-a	-2.4	-44.5	-47.5	-2.5	-59.3
MC-RR	7.9	5.0	7.4	3.7	3.0
Nodularin	4.0	3.5	5.9	3.8	6.6
MC-YR	-0.2	-1.8	2.5	2.5	9.4
MC-LR	2.7	-5.4	0.5	4.6	4.1
MC-LW	-20.6	-18	-29	-14.2	-19.6
MC-LF	-12.9	-14.9	-18	-13.1	-6.4

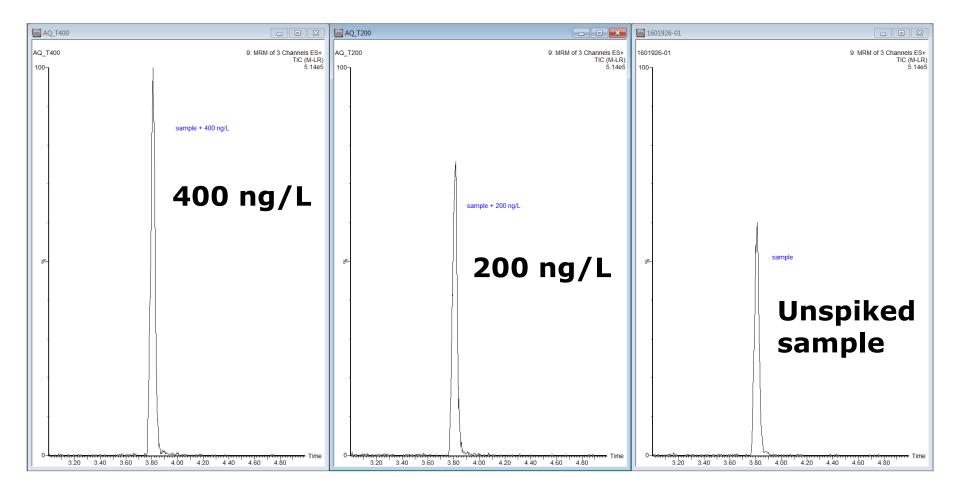
Results: Accuracy



- SW recoveries 30% 150% and dependent on type of surface water
- Suggests the need for IS to improve accuracy
- In the absence of an IS, standard addition can be used
- Standard addition calculations can be done automatically in TargetLynx
- Following slides show quan of M-LR in Aquacheck Proficiency Test using double addition. Assigned value for M-LR, based on 14 results, is 720 ng/L
- Sample analyzed using double standard addition:
 - Sample
 - Sample + 200 ng/L
 - Sample + 400 ng/L



Results: Standard Addition



Results: Standard Addition

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- In the Sample List:
 - Use "Standard" for sample type
 - Add the spiked concentration
 - Assign a sample group for each set of samples

	File Name	File Text	MS File	Inlet File	Bottle	Inject	Volume	Sample Type	Conc A	Sample Group
1	1601926-01	AQ500-groep S32	test_JD	Microcysteines A1-B1	1:10		50.000	Standard		aquacheck
2	AQ_T200	AQ_T200	test_JD	Microcysteines A1-B1	1:11		50.000	Standard	200	aquacheck
3	AQ_T400	AQ_T400	test_JD	Microcysteines A1-B1	1:12		50.000	Standard	400	aquacheck

- In the TargetLynx processing method:
 - Check the option "Use Standard Addition?"

C:\MassLynx projects\20160204 Watergroep microcystines.PRO\20160204 stadand addition.qld - TargetLynx XS Method Editor		8
File Edit Update View Compound Help		
D 2		
Compound List	2 2 2 2 4 4 4 5 2 2 2	
1: Anatoxin-a	Calibration Properties	Value
2: Nodularin	Compound Name	MC-LR
3: CYL		
4: MC-LF	Calibration Reference Compound	5: MC-LR
5: MC-LR		
6: MC-LW	Concentration Units	
7: MC-RR	Concentration of Standard: Level	Conc A
8: MC-YR	Stock Concentration Factor	⊠ 1.0000
9: MC-LY		
10: DE-MC-RR	Polynomial Type	Linear
	Calibration Origin	Exclude
	Weighting Function	1/X
	Ignore Zero Level Standards?	⊠ NO
	Innore Zoro Lever QUS?	
	Use Standard Addition?	♥ YES
	Propagate Calibration Parameters?	✓ YES

Results: Standard Addition



- TargetLynx calculates endogenous concentration
- Reported conc. 724 ng/L in good agreement with known value

File Edit View Display Processing Window Help	
	-LR
# Name Type Std. Conc RT Area Primary Flags Conc. //Dev Std. Add. Conc 1 1 1601926-01 Standard 0.000 3.81 7422.787 MM 6.3 724.3 2 2 AQ_T200 Standard 200.000 3.81 9262.519 bb 187.4 6.3 724.3 3 3 AQ_T400 Standard 400.000 3.81 11550.761 bb 412.6 3.2 724.3	
Calibration: 07 Apr 2016 15:42:03	Chromatogram
Compound name, MC-LR Correlation coefficient: r = 0.997523, r*2 = 0.995053	1601925-01 F9.MRM of 3 channels,ES+ A0500-groep S32 3.81 995.9 > 135.1
Calibration como 40 500 to v 7359 71	100 7 7423 1.776e+005
Curve type: Linear, Origin: Exclude, Weighting: 17c, as trans: None Standard Addition Concentration : 724.301	
300-j ×	
200	
1.00	
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g -1.00	
200 ·	
-3.00	
-400- 	
-5.00-	1601926-01 F9.MRM of 3 channels,ES+
	AQ500-groep S32 3.81 995.9 = 107.15 977 5458e+1004
x	
10000	
8000-	
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-0	0

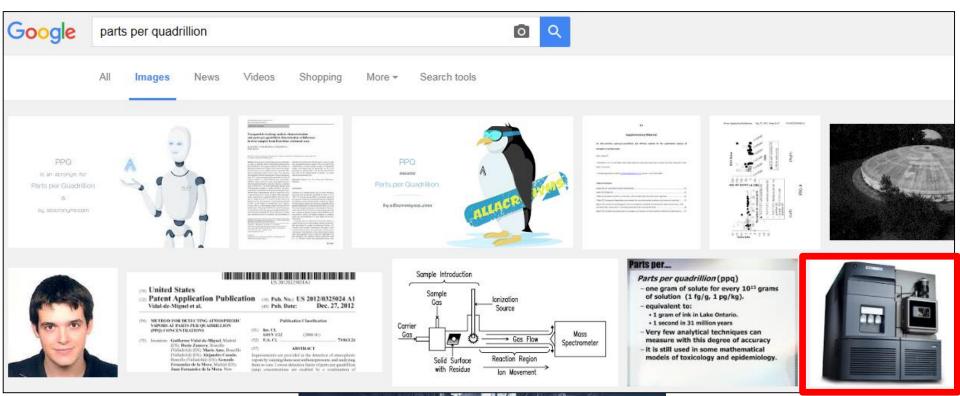
Summary of Direct Injection High Sensitivity TQ Method



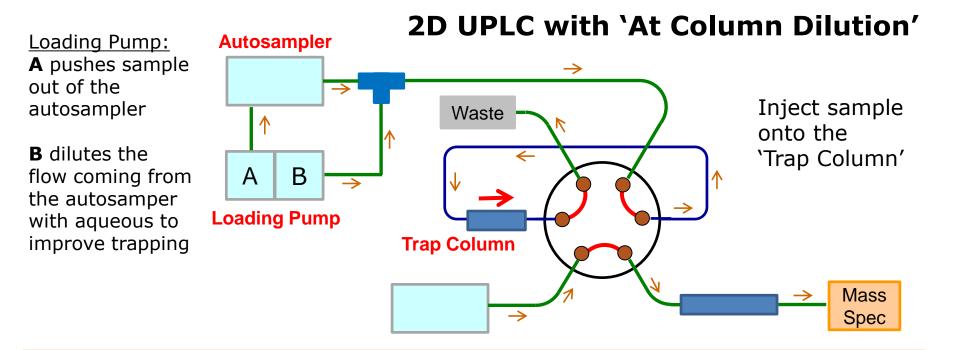
- Sensitive and robust method for 10 microcystins was developed:
 - 25 ng/L or lower LOD for all analytes
 - Signal stable for 15 hours during study
 - Repeatability <5% RSD
 - Linearity and %deviation excellent over the studied range
- Run time 7.5 min
- 50 μL of drinking water or surface water was injected directly
- In the absence of IS, standard addition yields accurate quantitation

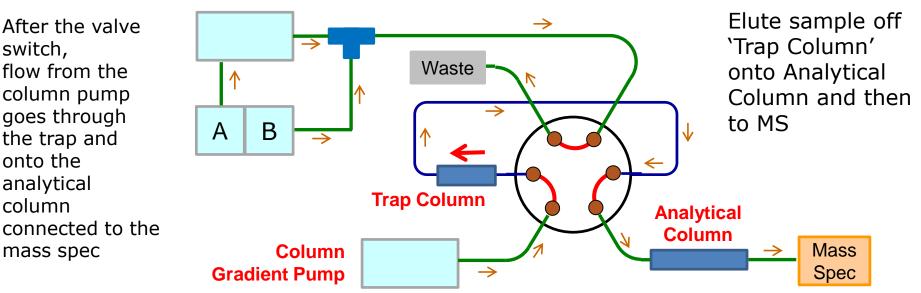
Beyond PPB and PPT: PPQ Detection













Experimental Parameters

ACQUITY I-Class 2D-UPLC ACD Method

Trap Column Oasis HLB Direct Conn 2.1 mm x 30 mm, 2(

Analytical Columi Waters ACQUITY UPLC E 130Å, 1.7 µm, 2.1 mm X 50

> UPLC System: ACQUITY I-Class Bir Solvent Manager (B ACQUITY Column Manage



250 µL Injections of processed water sample

Purge Solution = 90/10 H2O/MEOH

Wash Solution = 30/10 ACN/ IPA/MEOH/H2O

ple Temperature = 7 °C

Loading Pump

A = 67/33 Acetonitrile/Water					
B = Water					
A & B have 0.3% Formic Acid					

Gradient (Flow=0.80 mL/min)

Column Gradient Pump

 $\begin{array}{l} A = \mbox{Water with} \\ B = \mbox{Acetonitrile with} \\ A \& B \mbox{ have } 0.01\% \mbox{ Formic Acid} \end{array}$

Gradient (Flow=0.45 mL/min)

Time	<u>A%</u>	<u>B%</u>		Time	<u>A%</u>	<u>B%</u>	
0.0 min	15	85	– Switch at 2.25 –	0.0 min	90	10	
3.3 min	15	85		3.3 min	90	10	
3.4 min	95	5					
6.0 min	95	5					
6.5 min	15	85					
				7.3 min	5	95	
				8.5 min	5	95	
			- Switch at 10 25 -	9.0 min	90	10	
11.0 min	15	85	– Switch at 10.25 –				
11.5 min	15	85		11.5 min	90	10	
				c .			

Loading Pump:

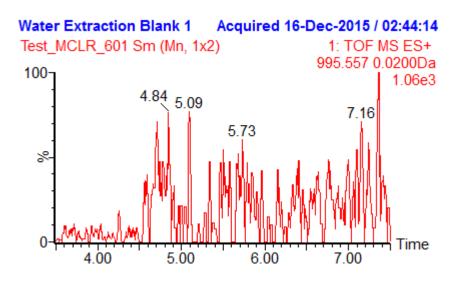
A is used to push the sample out of the autosampler

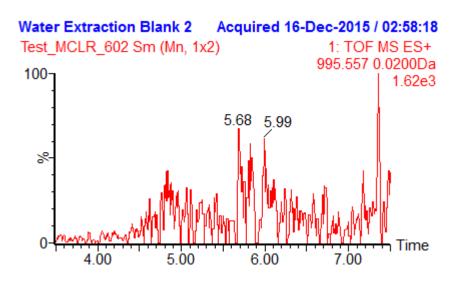
B is used to dilute the flow coming from the autosamper with aqueous before it gets to the trap column

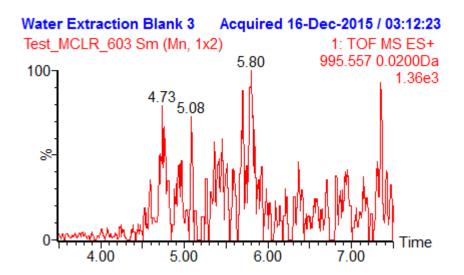
t=0: Flow from autosampler goes through trap column to waste t=2.25: valve switch to send flow from the column pump through the trap and onto the analytical column t=10.25 switch back to t=0 flow path

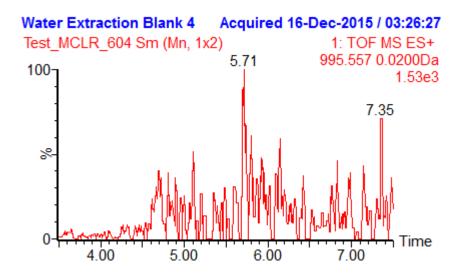


Blank Injections 1 - 4

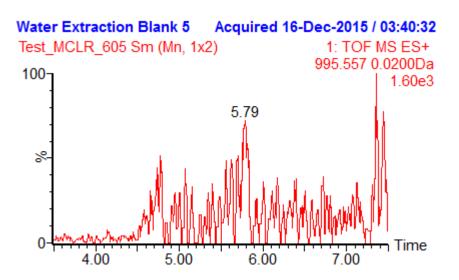




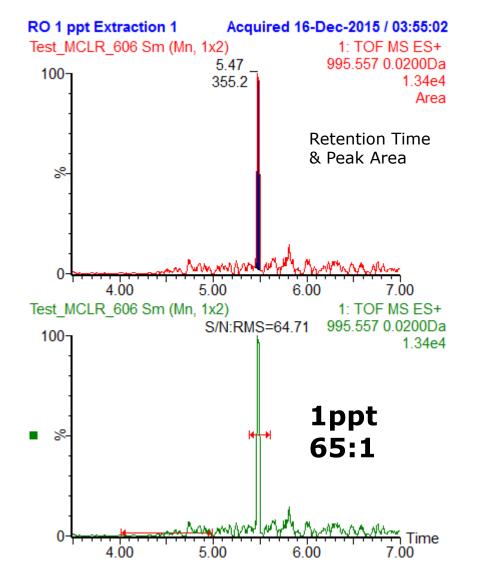




Blank Injection 5 and & RO Water Injection 1

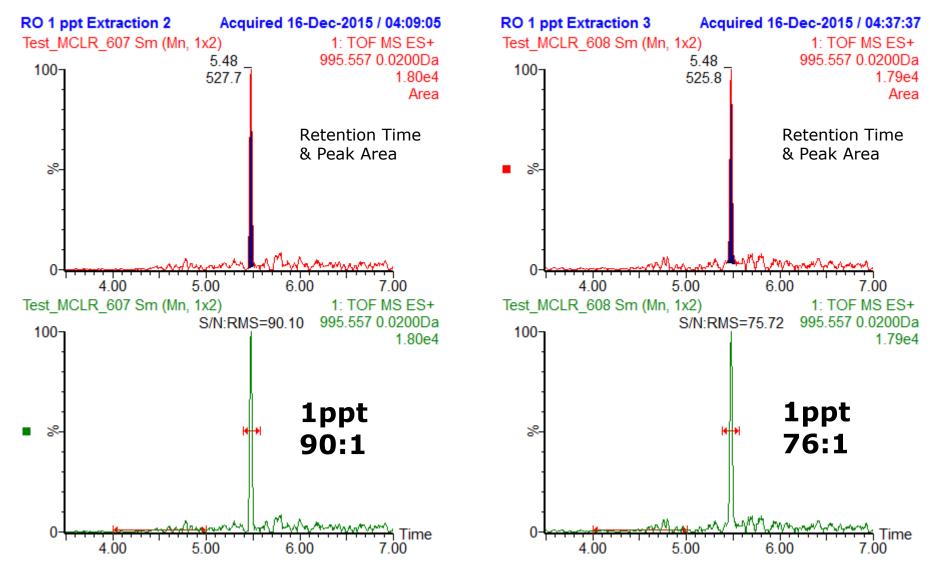






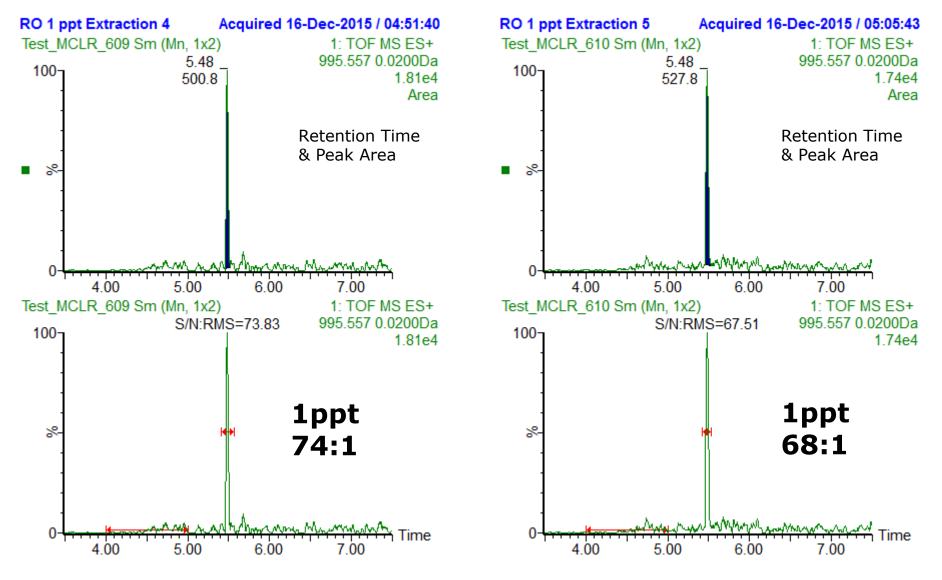


RO Water Injections 2 & 3





RO Water Injections 3 & 4



Instrument Detection Limits



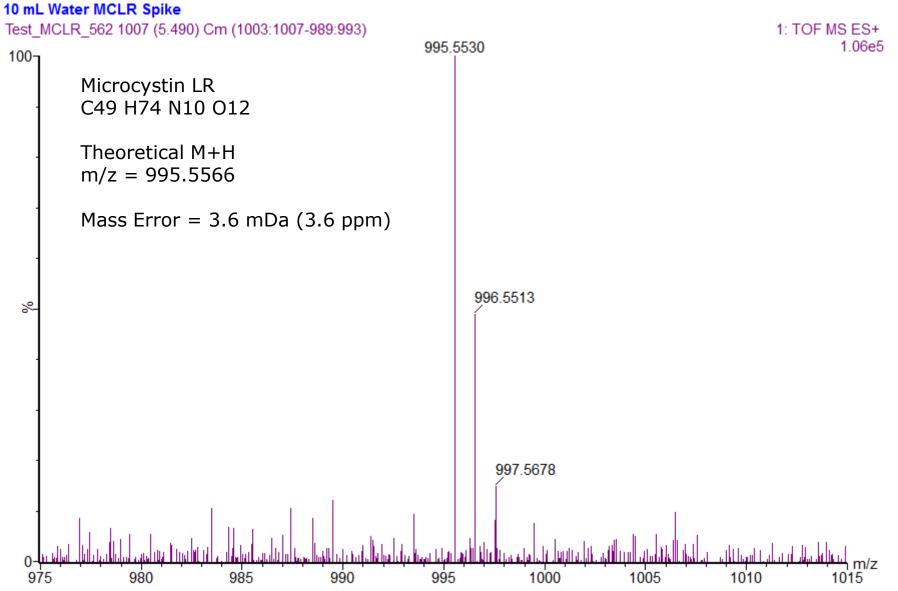
Five Analyses of RO Water Samples Spiked at 1 ppt

<u>Injection</u>	<u>RT</u>	<u>Area</u>	<u>RMS S:N</u>
1	5.48	355.2	64.71
2	5.48	527.7	90.10
3	5.48	525.8	75.72
4	5.48	500.8	73.83
5	5.48	527.8	67.51
	Average =	487.5	74.37
	Std Dev =	74.8	9.87
	RSD =	15.3%	13.3%

Translates to LOD of approximately <100ppq

Spectrum for 1 ppt of Microcystin LR

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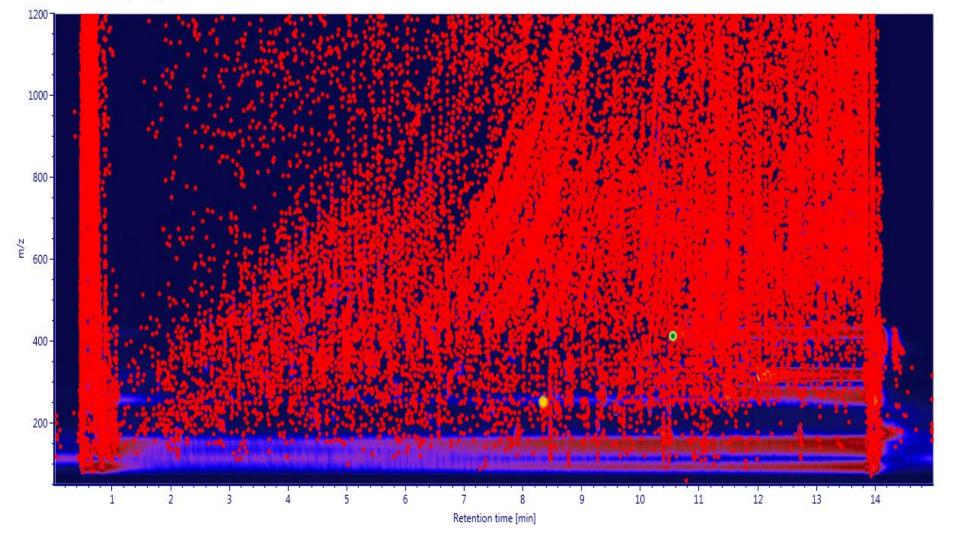
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Why Use a QTof?



Item name: Water_005 Channel name: 1: TOF MSe (50-1200) 6eV ESI+

Because you need to know more.

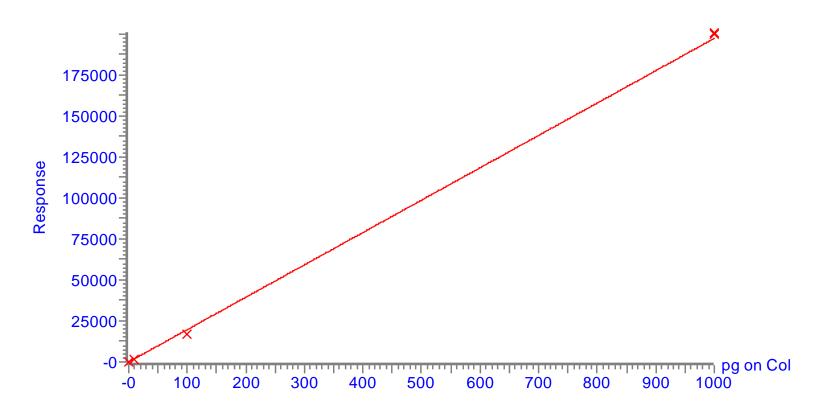


Linearity for Microcystin LR

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Compound name: Microcystin LR Correlation coefficient: r = 0.999008, $r^2 = 0.998017$ Calibration curve: 197.411 * x + -5.88617Response type: External Std, Area Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None

Calibration Curve for Microcystin LR 0.010ppb to 100ppb



Summary of 2D LC QTof Method



- Sensitive (PPQ) detection for MC-LR was achieved using 2D LC
 - 100 pg/L detection limit achieved
 - Linearity from 0.01 to 100ppb >0.99 R^2
 - Matrix effects were evaluated minimal and acceptable
- Run time <15 min</p>

Overall Summary

- Fit for purpose LOD can be achieved with high performance TQ through direct injection and little or no sample pretreatment
- 2D LC and SPE provide enrichment factors leading to fit for purpose LOD for lower sensitivity TQ or unparalleled sensitivity for research purposes when paired with high performance MS

Acknowledgements

De Watergroep, Belgium



Waters

Ministry of Environment and Climate Change, Toronto

